

# SMILE

Patient Safety Monitoring in International Laboratories



## **Our Mission:**

**To Provide Quality Assurance Support for  
DAIDS-Funded HIV Clinical Trial  
Laboratories (Non-US)**

**To Maintain a Web-Based Repository of  
Records and Resources**

Mark Swartz, MT(ASCP)

Anne Sholander, MT(ASCP)

International QA/QC Coordinators, SMILE

# ACKNOWLEDGEMENTS



DAIDS - Daniella Livnat and Mike Ussery

*This project has been funded in whole or in part with Federal funds from the Division of AIDS (DAIDS), National Institute of Allergy and Infectious Diseases, National Institutes of Health, Department of Health and Human Services, under contract No. HHSN266200500001C, titled Patient Safety Monitoring in International Laboratories.*



Johns Hopkins University – SMILE Staff

Dr. Robert Miller - Principal Investigator

Barbara Parsons - Operations Manager

Kurt Michael - Project Manager

# Who is SMILE?

- Contract resource between NIH/NIAID-DAIDS & Johns Hopkins University
- Staff of 12 with a wide range of laboratory experience

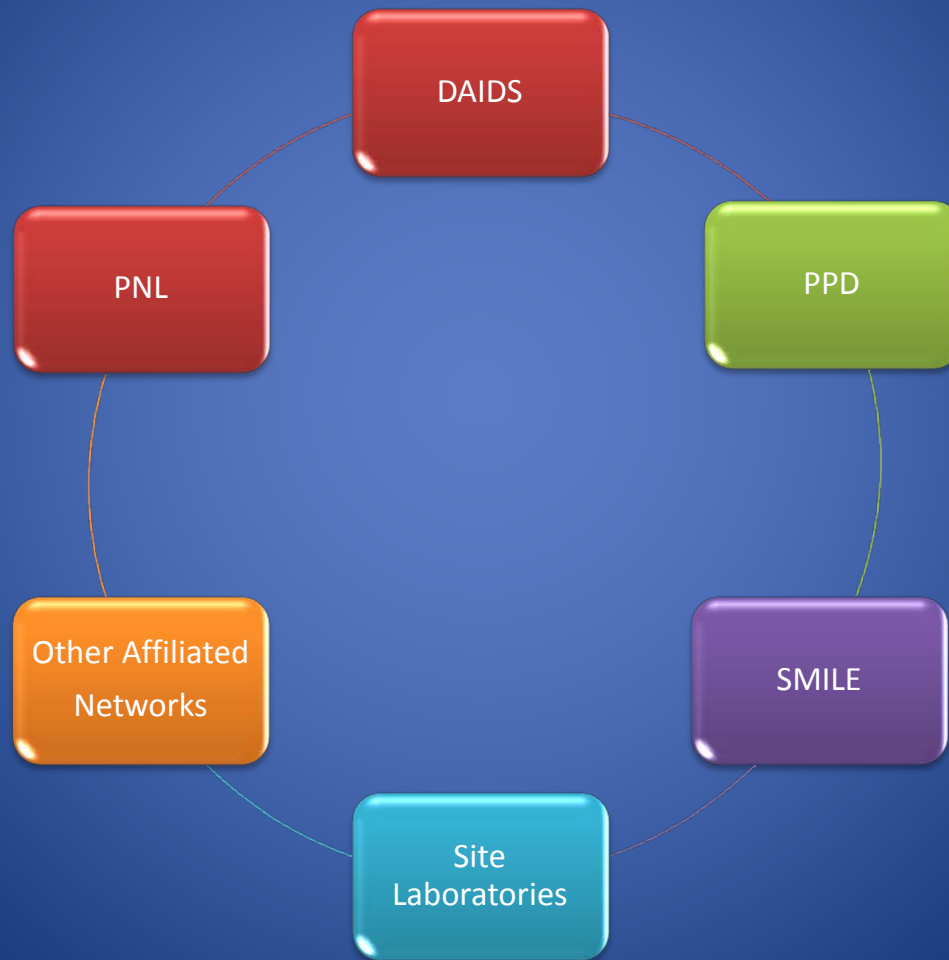


# Major Functions of SMILE

- Action Plans
- Validation Review
- Protocol Analyte Lists
  - EQA Review
  - Investigations
- Resources/Training



# Who are the partners?



# Laboratory Audits

Independent assessments are performed annually by PPD or DAIDS



Based on the standards of Good Clinical Laboratory Practice (GCLP)

**Clinical Research Support Contract No. N01-AI-50022**

*This project has been funded in whole or in part with Federal funds from the Division of AIDS (DAIDS), National Institute of Allergy and Infectious Diseases, National Institutes of Health, Department of Health and Human Services, under contract No. N01-AI-50022, titled HIV Clinical Research Support (CRS) Services.*



**Laboratory Audit Visit of**

**Conducted by PPD Laboratory Services**

Final Report Issued: 08 September 2011

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*To be completed on PPD Site File Copy ONLY.*

Audited By: \_\_\_\_\_ Signature: \_\_\_\_\_ Date: \_\_\_\_\_  
(Print Name)

Reviewed By: \_\_\_\_\_ Signature: \_\_\_\_\_ Date: \_\_\_\_\_  
(Print Name)

- Ordered by the DAIDS POC for your site
- Pre-visit checklist sent about 2 weeks before audit

## IX. Test and Control Articles, *continued*

### C. QC Materials

Comments:

3. QC materials were not dated with new expiration dates after opening or reconstitution; however, materials were labeled with the opened or preparation dates, and the stability of the materials was documented in the written procedures.

### D. Calibration Materials

1. Are calibration materials utilized by the laboratory? (If "No", skip to Section E.)	Yes <input checked="" type="checkbox"/>	No <input type="checkbox"/>	Comments <input type="checkbox"/>
2. Are all calibration materials dated within the manufacturer's assigned expiration dates?	Yes <input checked="" type="checkbox"/>	No <input type="checkbox"/>	Comments <input type="checkbox"/>
3. Are all calibration materials properly stored, as required by the manufacturer?	Yes <input checked="" type="checkbox"/>	No <input type="checkbox"/>	Comments <input type="checkbox"/>
4. Are all calibration materials properly labeled to indicate identity, lot number, storage requirement, date prepared/reconstituted, and expiration date?	Yes <input type="checkbox"/>	No <input checked="" type="checkbox"/>	Comments <input checked="" type="checkbox"/>

Comments:

4. Calibration materials were not dated with new expiration dates after opening or reconstitution; however, materials were labeled with the opened or preparation dates, and the stability of the materials was documented in the written procedures.

### E. Reagent/Testing Kits

1. Are all reagent/testing kits dated within the manufacturer's assigned expiration dates?	Yes <input checked="" type="checkbox"/>	No <input type="checkbox"/>	Comments <input type="checkbox"/>
2. Are all reagents/testing kits properly stored, as described by the manufacturer?	Yes <input checked="" type="checkbox"/>	No <input type="checkbox"/>	Comments <input type="checkbox"/>
3. Are all reagents/solutions properly labeled to indicate identity, lot number, storage requirement, date prepared/reconstituted, and expiration date?	Yes <input type="checkbox"/>	No <input checked="" type="checkbox"/>	Comments <input checked="" type="checkbox"/>

Comments:

3. Hematology reagents were not labeled with the new expiration dates after opening; however, the reagents were consumed well within the expiry dates, according to the Hematology Coordinator, Agda Moraes.



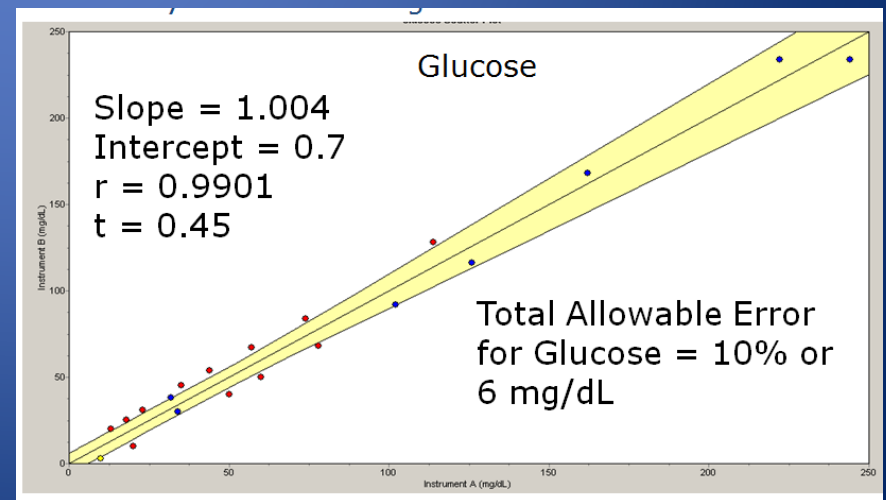
# Audits and Action Plans

1. Audit report sent to SMILE
2. Action Plan is prepared
3. SMILE returns the AP to DAIDS POC for distribution to the affiliated networks
4. Networks designate critical items
5. Action Plan and audit report sent to lab by DAIDS POC

A	B	C	D	E	F	G
1	Key			<p style="text-align: center;"><i>Brazil</i> HPTN, IMPAACT</p> <p style="text-align: center;"><i>PPD Audit Visit 15-21 December 2010</i></p> <p style="text-align: center;">Revised: 20 September 2011 by Anne Sholander</p>		
2	Complete					
3	In Progress					
4	Status Unknown		SMILE comments in BLUE.			
5	Contingent or Critical Items		Site comments in RED.			
6	Action Recommended; Follow-up Not Expected					
Audit SUB-SECTION	OBSERVATION / FINDING	SUGGESTED ACTION	HISTORY	REVIEWER COMMENTS	STATUS & DATE COMPLETED	
<b>SECTION II- ORGANIZATION AND PERSONNEL</b>						
1	II. A	<p><b>Organization chart--</b> a section specific organizational chart was not available for the Chemistry department. The job titles on the organizational charts were not consistent with the job titles listed in personnel records.</p>	<p>Create a section specific organogram for the Chemistry department. Review the organograms in all departments to ensure that the job titles listed on the organograms are consistent with the job descriptions in personnel records. Submit updated organograms to SMILE for review.</p>	<p><b>09 Mar 11:</b> Chemistry Organogram received. Revised organograms for Serology, Hematology and Flow Cytometry received. (AS)</p>	14-Mar-11	
2	II. B. 1-8 XIII. A. 1, 5	<p><b>Availability of personnel records--</b> the auditor noted that personnel records were not available for review because they were stored off site and the administrative office required one week's advance notice for retrieving the files.</p>	<p>For future audits, please make sure that all required records are available for review by the auditor. The pre-audit checklist should serve as a reminder to request all personnel records in advance. Notify SMILE when complete.</p>			
3	II. B. 1	<p><b>Personnel records--</b> no personnel file was maintained for the Section Coordinator in Serology. One Medical Coordinator personnel file contained no records.</p>	<p>Create a personnel file for the Section Coordinator in Serology. Update the personnel file for the Medical Coordinator with the appropriate records. Notify SMILE when complete.</p>			
4	II. B. 2	<p><b>Job descriptions--</b> two job descriptions had not been signed (laboratory technicians in Flow Cytometry and Chemistry).</p>	<p>Ensure that the two job descriptions cited have been signed by the appropriate personnel. Notify SMILE when complete.</p>	<p><b>20 Sept 11:</b> signed job descriptions submitted to SMILE. (AS)</p>	20-Sep-11	
5	II. B. 3	<p><b>Position profiles--</b> there was no job profile for the QA/QC Unit position.</p>	<p>Update the listing of position profiles to include the educational and experience requirements for the QA/QC Unit position. Submit the updated list to SMILE when complete.</p>	<p><b>31 Aug 11:</b> Updated position profiles received. (AS)</p>	31-Aug-11	
6	II. B. 5	<p><b>Assay specific training records--</b> training provided by the manufacturers was not documented in personnel files. Ortho provided training certificates by the end of the audit. Training for the Sysmex analyzer was not documented.</p>	<p>Ensure that training certificates are obtained from manufacturers performing assay specific training. Document all training in the appropriate personnel files. Notify SMILE when the Sysmex training certificate is received.</p>			
		<p><b>Competency records--</b></p>		<p><b>01 Aug 11:</b> The competency</p>		

# Validation Review and Assistance

- Common findings from PPD audits
- New protocols/new tests
- New instrumentation
- Request from networks
- Lab moving





- Home
- About pSmile
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- Resources
- People
- Organizations
- Contact Us

Quick Links

- ASLM Resource Center
- SMILE-PNL Investigation Form V4.0.
- SMILE-PNL Investigation Form for Microbiology V4.0.
- FDA-Approved Rapid HIV Ab Screening Test Chart
- CD4 and CD8 Method Validation Outline
- FDA Protocol Analyte List (PAL) - 16 Sep 2009
- EQA Providers - Other
- EQA Providers- Protocol Preferred

Welcome to Project pSMILE!

This web site is operated by the Johns Hopkins University for the pSMILE project (Patient Safety Monitoring & International Laboratory Evaluation) sponsored by the National Institutes of Health (NIH) Division of AIDS (DAIDS) under contract HHSN 266 2005 000 IC Miller (PI) 11/16/04-11/15/11.

The Resource section of pSMILE has been reorganized! Resources are now categorized by topic and the search capabilities have been expanded to make it easier for you to find what you need.

Where applicable, documents and links have references to the PPD audit within the description area. If you have any problems or questions, please contact your SMILE representative or use the [Contact Us](#) tab for your inquiries.

Please click [here](#) to request access to this web site.

# Types of validation resources

- Total Allowable Error (TEa) limits table
- Validation Overviews
- Detailed validation guidelines/instructions
  - Chemistry, Hematology, HIV EIA, Rapid HIV, Viral Markers EIA, Coagulation
- Statistical calculation tools
- Plan and Summary templates for each type of assay

# What are we looking for??

- Precision
- Accuracy
- Linearity (Analytical Measurement Range + Clinical Reportable Range)
- Sensitivity and Specificity
- Reference Ranges

### Quantitative Validation Overview

Validation of a quantitative system (for example Chemistry analyzer or Hematology analyzer) consists of an established set of required experiments. Each laboratory should first design a validation plan describing how they will satisfy each of these requirements. The validation plan must also detail the acceptability criteria for each element. After completing all of the validation experiments, results should be compiled and filed in an organized manner. All validation records should be retained for the life of the instrument. A validation summary should be prepared that contains a place for the Laboratory Director to sign, indicating the validation has been reviewed and approved.

The following are the required components of validation:

1. **Precision** is reproducibility - the agreement of the measurements of replicate runs of the same sample. Replication experiments are performed to estimate the imprecision or random error of the analytical method. See SMILE Precision Guidelines.
2. **Accuracy** is the true value of a substance being measured. Verification of accuracy is the process of determining that the test system is producing true, valid results. See SMILE Accuracy Guidelines.
3. **Linearity** - A quantitative analytical method is said to be linear when measured results from a series of sample solutions are directly proportional to the concentration or activity in the test specimens. This means that a straight line can be used to characterize the relationship between measured results and the concentrations or activity levels of an analyte for a determined range of analyte values. See SMILE Linearity Guidelines.
4. The **Analytical Measurement Range (AMR)** is the range of analyte values that a method can directly measure on the specimen without any dilution, concentration, or other pretreatment not part of the usual assay process. AMR validation is the process of confirming that the assay system will correctly recover the concentration or activity of the analyte over the AMR. The manufacturer defines the AMR – but it is the laboratory's responsibility to verify it. See SMILE AMR and CRR Guidelines.
5. The **Clinical Reportable Range (CRR)** is the range of analyte values that a method can report as a quantitative result, allowing for specimen dilution, concentration or other pretreatment used to extend the AMR. The laboratory should establish a CRR that covers the range of a Grade 4 Adverse Event on the DAIDS Toxicity Table

# For each of the required elements...



- Minimum number of samples tested
- Appropriate samples selected
- Comparison to a previously validated method
- Evaluation criteria clearly stated
- Evidence of comparison to manufacturer's specifications
- Raw data that supports conclusions



## SMILE Chemistry Precision Requirements

**PRECISION** is reproducibility - the agreement of the measurements of replicate runs of the same sample. Replication experiments are performed to estimate the imprecision or random error of the analytical method.

### I. Short-Term (Within Run/Day)

#### A. Sample:

1. Two levels (Low / High or Normal / Abnormal)
2. Patient or quality control

B. Testing: Run each sample 20 times on the same run, if possible, or at a minimum within the same day

#### C. Acceptability criteria:

1. Calculate the coefficient of variation (CV) for each level using 20 data points.
2. Compare to manufacturer's stated precision claims found in the package insert.
3. If manufacturer's precision cannot be met, it is acceptable to attain precision that is <25% of SMILE criteria for total error. Refer to the SMILE Precision Requirements Appendix 1 SMILE Chemistry TE Table.
4. If Short-Term precision is unacceptable, consult instrument manufacturer for assistance.

### II. Long-Term (Between Day)

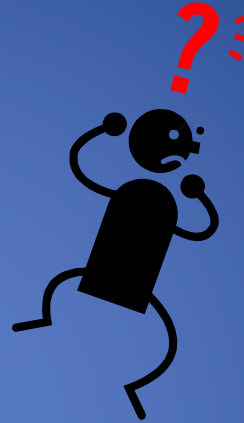
#### A. Sample:

1. Two levels (Low/High or Normal/Abnormal)
2. Patient or control serum. A lab may already have this data on hand from their daily QC runs.

B. Testing: Run each sample two times per day for 10 days for a minimum of 20 total data points for each level of material used.

#### C. Acceptability criteria:

# What is an auditor looking for??



- All of the above PLUS...
- All data summarized in an easy-to-understand, organized report
- Raw data that supports the conclusions
- **LABORATORY DIRECTOR SIGNATURE**

Insert Lab Header here  
Include full name and address of Lab

## Validation Summary Report

**Purpose:** Validation Re-Validation Other:

**Description of Equipment/Process:**

Equipment/Process: Insert full name of analyzer (ex: Cobas Integra 400+)

Serial Number: Insert instrument serial number

Location: Insert name of lab, city, state and country

Date: Insert date range of validation studies

FDA Approval Status: Approved Not approved

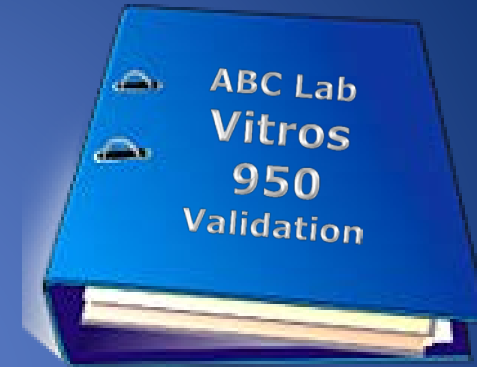
**Procedure:**

Refer to the insert lab name Validation Plan for insert instrument name  
Ex: Refer to the XYZ Lab Validation plan for Cobas Integra 400

**Results:** All raw data reports and statistical analysis can be found in the insert instrument name Validation binder.

1. Precision- refer to tab A

Analyte	Expected Results		Observed Results		Acceptability
	Manufacturer's Precision	33% of CLIA	Normal Control CV%	Abn Control CV%	
Ex:ALT	3.3%	6.6%	3.8%	4.3%	Acceptable



**Reference Range Approval**

Medical Director: \_\_\_\_\_ Date: \_\_\_\_\_  
Insert Medical director name here

**Method Approval**

Approved / Not Approved

If not approved, provide recommendations/corrective actions below.

Laboratory Director: \_\_\_\_\_ Date: \_\_\_\_\_  
Insert Lab director name here

Prepared by: \_\_\_\_\_ Date: \_\_\_\_\_  
Insert name and title here

# Protocol Analyte List (PAL)

- Protocol analytes
- Instrumentation in use
- FDA Approval status
- Back-up plans
- Specimen flow



**Laboratory Name: 7 CLINICAL TRIALS ON SITE LABS & XXX CENTRAL ROUTINE LAB & ABC/QRS DURBAN & JHB**  
**Protocol: MTN003**

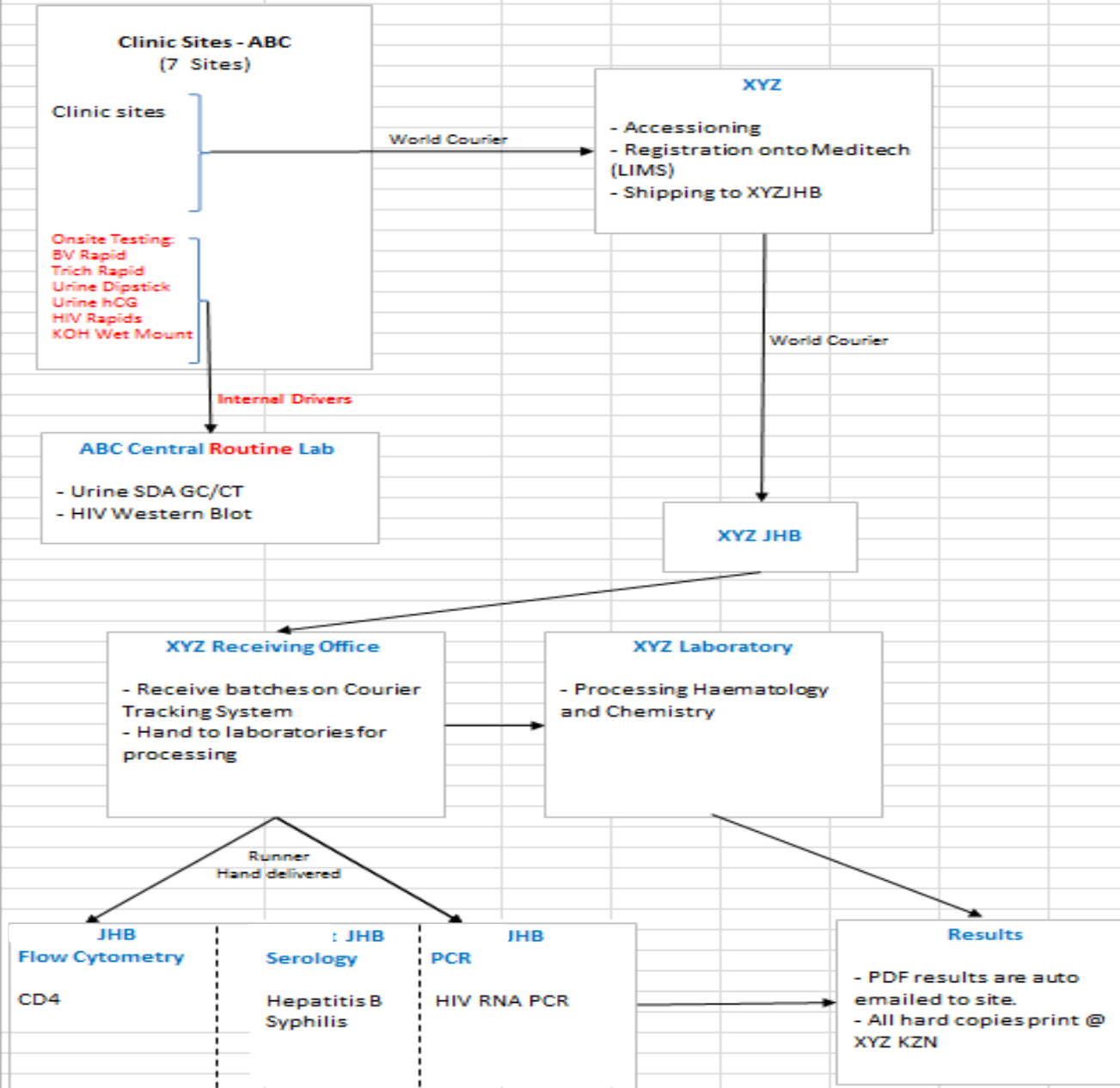
Completed By: \_\_\_\_\_  
 Date Completed: **9-May-11**

*Please fill in as much detailed information as you can provide. Please add any assays that are missing.*

Assay	Primary Instrument			Primary Method/Reagent Kit								
	Instrument Name	Instrument Manufacturer	Instrument Model Number	FDA Approved (Yes/No/D on't Know)	Method/Kit Name	Method/Kit Manufacturer	Method/Kit Product Number or Product Code	FDA Approved (Yes/No/D on't Know)	Primary laboratory performing testing for this	External QA provider	Frequency of EQA	EQA Panel Details
<b>Chemistry</b>												
ALT (SGPT)	COBAS INTEGRA	ROCHE	400 PLUS	FDA/CE	(IFCC), without pyridoxal-5'-phosphate	ROCHE	20764957	FDA/CE	ABC Lab JHB	CAP & THISTLE	3 per yr / 1 every two wks	CAP C3 X 5 MEMBER
AST (SGOT)	COBAS INTEGRA	ROCHE	400 PLUS	FDACE	(IFCC), without pyridoxal-5'-phosphate	ROCHE	20764949	FDA/CE	ABC Lab JHB	CAP & THISTLE	3 per yr / 1 every two wks	CAP C3 X 5 MEMBER
Phosphate (inorganic)	COBAS INTEGRA	ROCHE	400 PLUS	FDA/CE	Phosphomolybdate UV /Integra 400plus / Roche	ROCHE	3183793	FDA/CE	ABC Lab JHB	CAP & THISTLE	3 per yr / 1 every two wks	CAP C3 X 5 MEMBER
Creatinine	COBAS INTEGRA	ROCHE	400 PLUS	FDA/CE	Jaffé Compensated	ROCHE	20764345	FDA/CE	ABC Lab JHB	CAP & THISTLE	3 per yr / 1 every two wks	CAP C3 X 5 MEMBER
<b>Urinalysis</b>												
Dipstick Urinalysis	N/A	N/A	N/A	N/A	Multi-stix 9, Uristix 4 and 2	Siemens	N/A	YES	XYZ Clinics	CAP	Twice Annually	CMM
Urine β-hCG	N/A	N/A	N/A	N/A	Quidel Quickvue	Quidel	BMX/97016	YES	XYZ Clinics	CAP	Twice Annually	CMM
<b>Hematology</b>												
Hemoglobin	COULTER	BECKMAN COULTER	LH 750	FDA/CE	Coulter	BECKMAN COULTER	NA	FDA/CE	ABC Lab JHB	CAP	3 per yr	FH 6
Hematocrit	COULTER	BECKMAN COULTER	LH 750	FDA/CE	calculation	BECKMAN COULTER	NA	FDA/CE	ABC Lab JHB	CAP	3 per yr	FH 6
White Blood Cell (WBC) Count	COULTER	BECKMAN COULTER	LH 750	FDA/CE	Coulter principle	BECKMAN COULTER	NA	FDA/CE	ABC Lab JHB	CAP	3 per yr	FH 6
Red Blood Cell (RBC) Count	COULTER	BECKMAN COULTER	LH 750	FDA/CE	Coulter principle	BECKMAN COULTER	NA	FDA/CE	ABC Lab JHB	CAP	3 per yr	FH 6
Platelet Count	COULTER	BECKMAN COULTER	LH 750	FDA/CE	Coulter principle	BECKMAN COULTER	NA	FDA/CE	ABC Lab JHB	CAP	3 per yr	FH 6
WBC Differential Count	COULTER	BECKMAN COULTER	LH 750	FDA/CE	VCS	BECKMAN COULTER	NA	FDA/CE	ABC Lab JHB	CAP	3 per yr	FH 6
<b>Bacteriology</b>												
GC/CT	Probetec	BD	Probetec1 - PT-2402 / Probetec-2 - PT-2167	Yes	SDA	BD	Without AC-440705 ,With AC-440450, Controls-440451, Diluent-	Yes	XYZ Routine Lab	CAP	Thrice Annually	HC6
<b>Point of Care Testing</b>												
BV Rapid	N/A	N/A	N/A	N/A	OSOM Rapid BV	OSOM/GRYPHUS	BVB25-1	Yes	XYZ Clinics	None	None	N/A
Trich Rapid	N/A	N/A	N/A	N/A	OSOM Rapid Trich	OSOM	181E	Yes	XYZ Clinics	CAP	Thrice Annually	VS1
KOH Wet Mount	N/A	N/A	N/A	N/A	Manual KOH preparation	N/A	N/A	N/A	XYZ Clinics	CAP	Thrice Annually	CMM

**Specimen Flow Chart for : ABC**

*Completed by:* \_\_\_\_\_  
*Date Completed:* **2010/10/14 - Updated 3 May 2011**



# Proficiency Testing (EQA)



College of American Pathologists



Proficiency Testing (PT) is a means of evaluating a laboratory's performance through analysis of unknown samples provided by an external source.



# EQA Summary Schedule

	A	B	C	D	E	F	G	H	I
1									
2									
3									Investigation Report Required
4		2009				Lab Name			Accutest# CAP#:
5		Ship Date	EQA Provider	Survey Sequence	Module Code	Test Group Name	SMILE Reviewed (Date)	Site Response (Date)	Comment
7	1	5-Jan	CAP	A	VM-1	Viral Markers	5-Mar-09	N/A	100% Successful
8	2	19-Jan	CAP	A	P	Parasitology	5-Mar-09	N/A	100% Successful
9	3	26-Jan	CAP	A	BP	Blood Parasites	1-Apr-09	8-May-09	Investigation required
10	4	26-Jan	CAP	A	FH3	Hematology w/Auto Diff and Photos	1-Apr-09	30-Apr-09	Investigation required for monocytes, neutrophils and blood cell ID.
11	5	9-Feb	CAP	A	E1	Limited Mycobacteriology	5-Jun-09	N/A	100% Successful
12	6	23-Feb	CAP	A	D5	Gram Stain	7-May-09	N/A	100% Successful
13	7	2-Mar	CAP	A	C3	General Chemistry	5-Jun-09	N/A	100% successful
14	8	16-Mar	CAP	A	CM	Clinical Microscopy	7-May-09	N/A	100% Successful
15	9	30-Mar	CAP	A	D4	Bacteriology	5-Jun-09	N/A	100% Successful
16	10	30-Mar	CAP	A	AHIV	Raphid HIV-1/2	5-Jun-09	N/A	100% Successful
17	11	13-Apr	CAP	A	G	Syphilis Serology	5-Jun-09	N/A	100% Successful
18	12	4-May	CAP	B	FH3	Hematology w/Auto Diff and Photos	9-Jul-09	6-Aug-09	Investigation for Cell ID
19	13	4-May	CAP	B	P	Parasitology	9-Jul-09	18-Aug-09	Investigation for Parasite ID and Photopages ID
20	14	11-May	CAP	B	VM-1	Viral Markers	9-Jul-09	NA	100% Successful
21	15	11-May	CAP	A	XU6	Serum HCG	5-Jun-09	N/A	100% Successful
22	16	18-May	CAP	B	BP	Blood Parasites	4-Aug-09	27-Aug-09	Investigation received for Blood Parasite ID, accepted
23	17	1-Jun	Accutest	2	BACT435	Bacteriology	11-Sep-09	N/A	100% Successful
24	18	1-Jun	Accutest	2	GRAM435	Gram Stain	11-Sep-09		Investigation required for gram stain.
25	19	1-Jun	Accutest	2	HEFD435	Hematology	4-Aug-09	N/A	100% Successful
26	20	1-Jun	Accutest	2	CELL435	Blood Cell ID	4-Aug-09	08/27/09	Investigation received, accepted
27	21	1-Jun	Accutest	2	SHCG435	Serum HCG	4-Aug-09	N/A	100% Successful
28	22	1-Jun	Accutest	2	BCHE435	Chemistry	4-Aug-09	N/A	100% Successful
29	23	1-Jun	Accutest	2	HIV435	Raphid HIV-1/2	4-Aug-09	N/A	100% Successful
30	24	1-Jun	Accutest	2	SYPH435	Syphilis Serology	4-Aug-09	N/A	100% Successful
31	25	1-Jun	CAP	B	C3	General Chemistry	4-Aug-09	N/A	100% Successful
32	26	6-Jul	CAP	B	D5	Gram Stain	11-Sep-09	N/A	100% Successful
33	27	3-Aug	CAP	B	CM	Clinical Microscopy			
34	28	3-Aug	CAP	B	E	Mycobacteriology			
35	29	10-Aug	CAP	B	XS4	Serum HCG			
36	30	10-Aug	CAP	B	AHIV	Raphid HIV-1/2			
37	31	10-Aug	CAP	B	D4	Bacteriology			
38	32	10-Aug	CAP	B	G	Syphilis Serology			
39	33	24-Aug	CAP	C	VM-1	Viral Markers			
40	34	14-Sep	CAP	B	F1	Fungal			
41	35	21-Sep	CAP	C	FH3	Hematology w/Auto Diff and Photos			
42	36	21-Sep	CAP	C	BP	Blood Parasites			



	A	B	C	D	E	F	G	H	I	J	K	L	M
1	Protocol/Non Protocol Analytes: No Event or Not Ordered												
2	Non-protocol Only: Result Not Submitted or Unable to Evaluate												
3	Internal Investigation (INT) Recommended												
4	Investigation Report Required (<100% but ≥80%) due within 30												
5	Investigation Report (IR) Required (<80%) due within 30 days												
6	Non-protocol Analyte												
7	ACC - Acceptable												
8													
9													
10	<b>Survey</b>		<b>2007</b>			<b>2008</b>			<b>2009</b>				
			<b>A / 1</b>	<b>B / 2</b>	<b>C / 3</b>	<b>A / 1</b>	<b>B / 2</b>	<b>C / 3</b>	<b>A / 1</b>	<b>B</b>	<b>2</b>	<b>C / 3</b>	
11	<b>Chemistry</b>	ALT				100%	100%	100%	100%	100%	100%		
12		Albumin				100%	100%	80%	100%	100%	100%		
13	Vitros 250	Alk Phos				100%	100%	100%	100%	100%	100%		
14		AST				100%	100%	100%	100%	100%	100%		
15		Bili, Direct				100%	100%	100%	100%	100%	100%		
16		Bili, Total				100%	100%	100%	100%	100%	100%		
17		Calcium				100%	100%	100%	100%	100%	100%		
18		Potassium - K				100%	100%	100%	100%	100%	100%		
19		Total Protein				100%	100%	100%	100%	100%	100%		
20		Sodium - Na				100%	60%	100%	100%	100%	100%		
21		Phosphorus				100%	100%	100%	100%	100%	100%		
22	<b>Hematology</b>	WBC	100%	100%	100%			100%	100%	100%	100%		
23		RBC	100%	100%	100%			100%	100%	100%	100%		
24	CellDyn3200	Hgb	100%	100%	100%			100%	100%	100%	100%		
25		Hct	100%	100%	100%			100%	100%	100%	100%		
26	Primary Instrument	MCV	100%	100%	100%			100%	100%	100%	100%		
27		RDW	100%	100%	100%			80%	100%	100%	100%		
28		Platelet	100%	100%	100%			60%	100%	100%	100%		
29		Neutrophil	100%	100%	100%			100%	80%	100%	100%		
30		Lymphs	100%	100%	100%			100%	100%	100%	100%		
31		Eos	80%	100%	100%			100%	100%	100%	100%		
32		Basos	100%	100%	100%			100%	100%	100%	100%		
33		Mono	100%	100%	100%			100%	80%	100%	100%		
34		Blood Cell ID				100%	78%	90%	90%	90%	80%		
35	<b>Syphilis</b>	MHA-TP	100%	100%	100%	100%	100%	100%	100%		100%		
36		RPR	100%	100%	80%	100%	100%	100%	100%		100%		
37		RPR Titer	100%	100%	80%	60%	100%	80%	100%				
38	<b>GC-Chlamydia</b>	Chlamydia	TNP	33%	66%								
39		GC	TNP	0%	100%								
40	<b>HCG</b>	Serum hCG (S)	100%	100%	100%		100%						
41		Serum hCG (XS-4)					100%	100%	100%			100%	
42	<b>Anti-HIV</b>	Method 1 (Abbott)	100%	100%	100%	100%	100%	100%	100%			100%	
43		Method 2 (Trinity)	100%	100%	40%	100%	100%	100%	100%			100%	
44	<b>Viral Markers</b>	HBsAg	100%	100%	100%	40%	100%	100%	100%	100%			
45		HBsAg Neutralz	100%	100%	100%	40%	100%	100%	100%	100%			

Lab Name: XYZ Lab  
 CAP# :  
 Accutest#:

# EQA Review

## SMILE – EQA Evaluation

Date: 12 July 2011

EQA Provider: CAP#

Site: ...

Panel: Clinical Microscopy, C-B 2011

### **Summary:**

This EQA event was successful for all protocol analytes except as noted below.

A table containing manually evaluated results can be found on the following page.

**Investigation Reports** are required for all **protocol analytes** scoring less than 100%.

The analytes requiring investigation are:

- None

**Internal investigations** are recommended for non-protocol analytes scoring less than 100% and for any bias, shifts and/or trends identified below:

Bias noted

Positive: None

Negative: AST, Creatinine, Sodium

Shifts noted: NA

Trends noted: NA

---

Non-Protocol Analytes scoring less than 100%:

- Sodium

# Manual Evaluation

EVALUATION ORIGINAL		VM-C 2010 Viral Markers		
Test Method	Specimen	Your Result	Intended Response	Your Grade
HIV-1 WB Proteins	VM1-27	NO MAJOR PROT DETECTED		[26]
	VM1-28	gp41		[26]
		p17/18		
		p24		
		p31/32		
		p51/55		
	VM1-29	p65/66		
		gp120/160		
		gp41		[26]
		p17/18		
		p24		
	VM1-30	p31/32		
		p51/55		
p65/66				
gp120/160				
gp41			[26]	
VM1-31	NO MAJOR PROT DETECTED		[26]	

## SMILE – Proficiency Evaluation

Date: 8 October 2010

Site:

Proficiency Provider: CAP #i

Panel: Viral Markers VM-C, 2010

### Summary:

This EQA event was successful for all protocol analytes.

The table contains manually evaluated results.

SAMPLE #	HIV-1 Western Blot		GRADE	HIV-1 WB Proteins		GRADE
	YOUR LAB RESULT	PEER RESULT		YOUR LAB RESULT	PEER RESULT	
VM1-27	Negative	Negative	Acceptable	Not Detected	Not Detected	100%
VM1-28	Positive	Positive	Acceptable	gp41 p24 p31/32 p51/55 p65/66 gp120/160	gp41 p24 p31/32 p51/55 p65/66 gp120/160	100%
VM1-29	Positive	Positive	Acceptable	gp41 p24 p31/32 p51/55 p65/66 gp120/160	gp41 p24 p31/32 p51/55 p65/66 gp120/160	100%
VM1-30	Positive	Positive	Acceptable	gp41 p24 p31/32 p51/55 p65/66 gp120/160	gp41 p24 p31/32 p51/55 p65/66 gp120/160	100%
VM1-31	Negative	Negative	Acceptable	Not Detected	Not Detected	100%
Average						100%

**Investigation Reports** are required for all **protocol analytes** scoring less than 100%.

The analytes requiring investigation are:

- None

## Summary:

This EQA event was successful for all protocol analytes except as noted below. Since no method was chosen CAP did not grade the survey. SMILE manually graded the survey; see survey for results.

The table contains manually evaluated results.

SAMPLE #	YOUR LAB RESULT	PEER RESULT	GRADE
CMMP-31	Sperm present	Sperm present	Acceptable

**Investigation Reports** are required for all **protocol analytes** scoring less than 100%. The analytes requiring investigation are:

- None

**Internal investigations** are recommended for the following non-protocol analytes scoring less than 100%:

- pH-33%
- Urobilinogen – 66%

## Comments:

Congratulations on your good EQA results.

Internal investigation is suggested for the pH and urobilinogen analytes.

On future surveys please enter the method code so CAP can grade the survey.

If you have any questions about your review please contact me.

Please remember to include both the method (Siemens) and instrument (Visual) on the next survey so CAP can grade this survey.

On the CAP instructions it specifies that when resulting of secondary confirmation methods (MHA-TP) where your process is to confirm only weakly reactive or reactive specimens **that leaving presumed nonreactive specimens blank will results in a penalty. Please enter a code for your non-reactive test on your confirmatory test in the future.**

# SMILE Investigation Form

## SMILE –Investigation Form

Date: 12-Jan-09		Site:	
Proficiency Provider & Panel: CAP C-C 2008			
Test Analyte (One/form if problems are unrelated):	Reported Result:	Acceptable Result:	Previous Survey Problems:
Albumin	4.3	3.3 – 4.2	No

**Laboratory Investigation:** (Attach supporting documentation. Please add detailed comments. Comment section will expand as needed.)

1. Survey report examined for discrepancies, clerical errors and appropriate codes?

Comments: **NA**

2. Survey material receiving temperature, handling, reconstitution, storage and analysis investigated?

Comments: **Received warm**

3. Method & instrument history reviewed (Daily preventative maintenance performed? Reagents –open date, expiration date, number of tests left on the reagent, instrument settings, etc.)?

Comments: **Yes**

4. Quality Control reviewed (Circle or mark appropriate answer below.)? **Yes**

Mean established by? parallel testing or manufacturer's used      QC ranges are set? Manufacture's range or locally established

QC within +/- 2 SD on day of survey? Yes No      Number of levels of QC run? 2 3

What QC rules do you use? **1\_3S**      Does your laboratory track Coefficient of Variation (CV) for analytes? CV = SD/Mean x 100 No

Comments: **QC was within limits (slightly on the high side) but there was a humidity issue that has been resolved since**

# Root cause analysis

## SMILE –Investigation Form

9. Personnel competency reviewed? Conduct staff education or retraining if applicable.

Comments: NA

10. Assess patient results for adverse impact? Review impacted patient results, amend results and notify physicians, if applicable.

Comments: The Vitros 250 weren't used yet to assay patient results

### Deficiency Classification:

- Methodological  
 Technical  
 Clerical

- Survey evaluation problem  
 Other (explain)

Summary of Actions: Briefly discuss what actions were taken in this investigation and what you believe is the primary cause of this EQA problem.

The problem was caused by an influx of humidity in the slide reagent chamber due to screws that were not secured tightly enough

Summary of Actions: Briefly discuss what has been done to prevent this problem from occurring in the future.

The problem was fixed and now the ) biomedical engineers will use this as a lesson in case similar a problem in the future

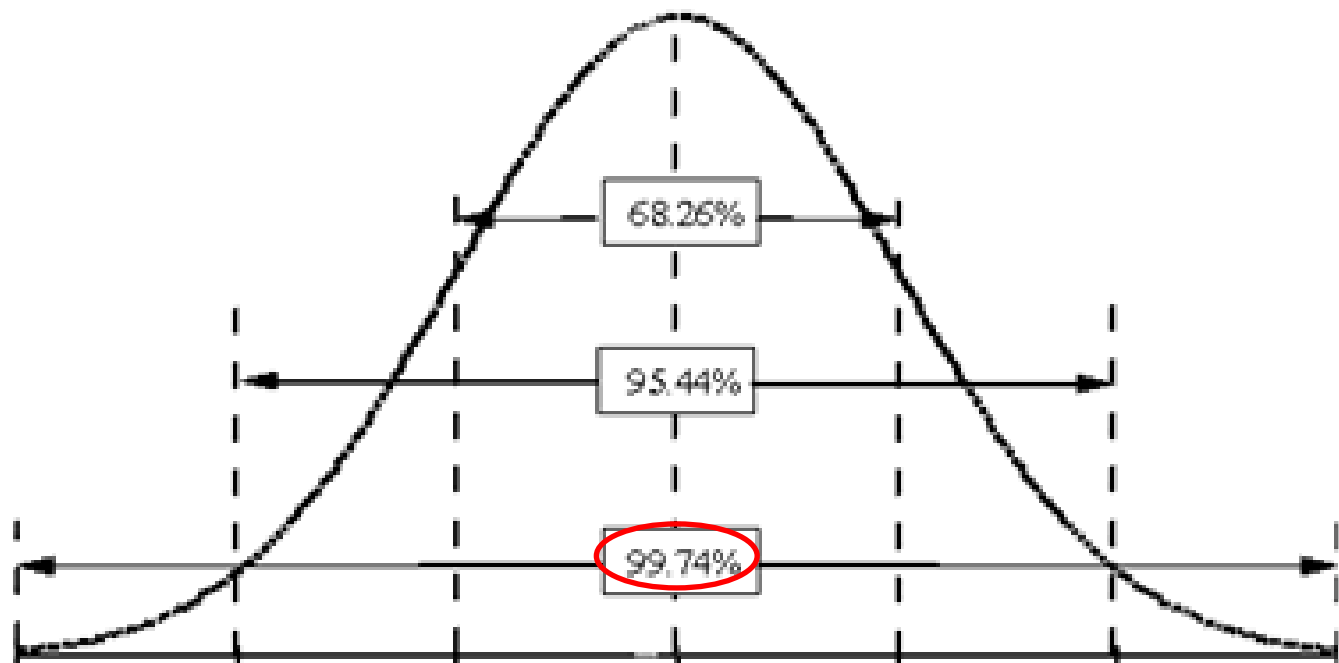
Prepared by:

Name/TITLE:

i, Lab QA Manager

Date: 14-Jan-09

Signature: MAG



$\bar{x} - 3s$        $\bar{x} - 2s$        $\bar{x} - 1s$        $\bar{x}$        $\bar{x} + 1s$        $\bar{x} + 2s$        $\bar{x} + 3s$

$14 - 3(2) = 8$        $14 - 2(2) = 10$        $14 - 1(2) = 12$       14       $14 + 1(2) = 16$        $14 + 2(2) = 18$        $14 + 3(2) = 20$



# Document Repository

EQA surveys, reviews and investigations on  
**[www.psmile.org](http://www.psmile.org)**

- Available to site labs (privately by log-in)
- Available to DAIDS and the Networks


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- Rapid HIV Testing Provider
- Network Protocol Analytes - 24 Sep2007
- SMILE-PNL Investigation Form V4.0
- SMILE-PNL Investigation Form for Microbiology V4.0

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**The Resource section of pSMILE has been reorganized! Resources are now categorized by topic and the search capabilities have been expanded to make it easier for you to find what you need.**

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#### Proficiency Testing Records

Instituto Dermatologico Y Cirugia de Piel(Dominican Republic)

	IDCP EQA Summary	10/05/2009	<a href="#">Update</a>
	IDCP CAP2009 Clinical Microscopy CM-B Review	10/04/2009	<a href="#">Update</a>
	IDCP CAP2009 Clinical Microscopy CM-B Survey	10/03/2009	<a href="#">Update</a>
	IDCP CAP2009 Chemistry C-B Review	08/04/2009	<a href="#">Update</a>
	IDCP CAP2009 Chemistry C-B Survey	08/03/2009	<a href="#">Update</a>
	IDCP CAP2009 Viral Markers VM-B Review	08/02/2009	<a href="#">Update</a>
	IDCP CAP2009 Viral Markers VM-B Survey	08/01/2009	<a href="#">Update</a>
	IDCP CAP2009 Hematology FH3-B Investigation	06/28/2009	<a href="#">Update</a>
	IDCP CAP2009 Hematology FH3-B Review	06/28/2009	<a href="#">Update</a>
	IDCP CAP2009 Hematology FH3-B Survey	06/27/2009	<a href="#">Update</a>
	IDCP CAP2009 Chemistry C-A Review	05/30/2009	<a href="#">Update</a>

# Document Repository

Also on [www.psmile.org](http://www.psmile.org):

- Audits (DAIDS and PPD)
- Action Plans
- Protocols and Protocol Analyte Lists (PAL)
- Multitudes of other resources


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# Major Functions of SMILE

- Action Plans
- Validation Review
- Protocol Analyte Lists
  - EQA Review
  - Investigations
- Resources/Training



# QUESTIONS?

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